



Sensitive Resistance Testing

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MTN Regional Meeting 2015

Lab Breakout Session

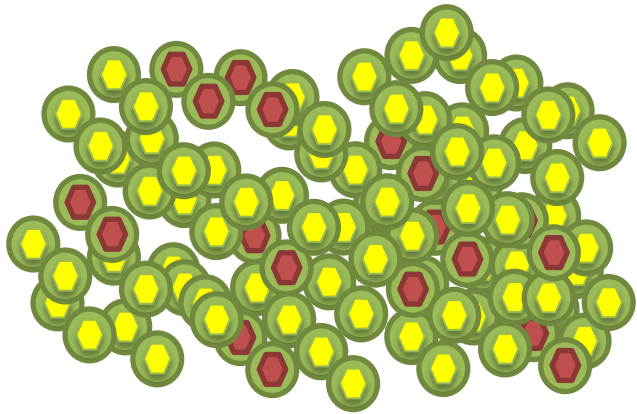
Outline

- Standard vs sensitive resistance testing
- Why does sensitive resistance testing matter?
- Resistance testing at the Virology Core
- Comparison of resistance tests
 - Standard (IHG) vs Sensitive (ASPCR and NGS)
- Sensitive resistance testing in VOICE and ASPIRE

Standard vs. Sensitive Resistance Testing

Standard

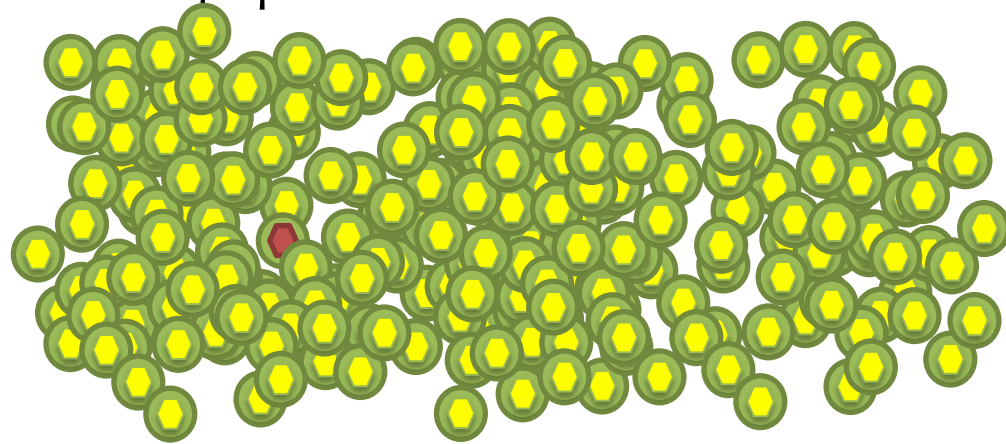
- Can detect drug resistance at a limit 20% of a patient's HIV virus population



- Results available in 1 week
- Can be used for clinical care
- Moderate cost per sample

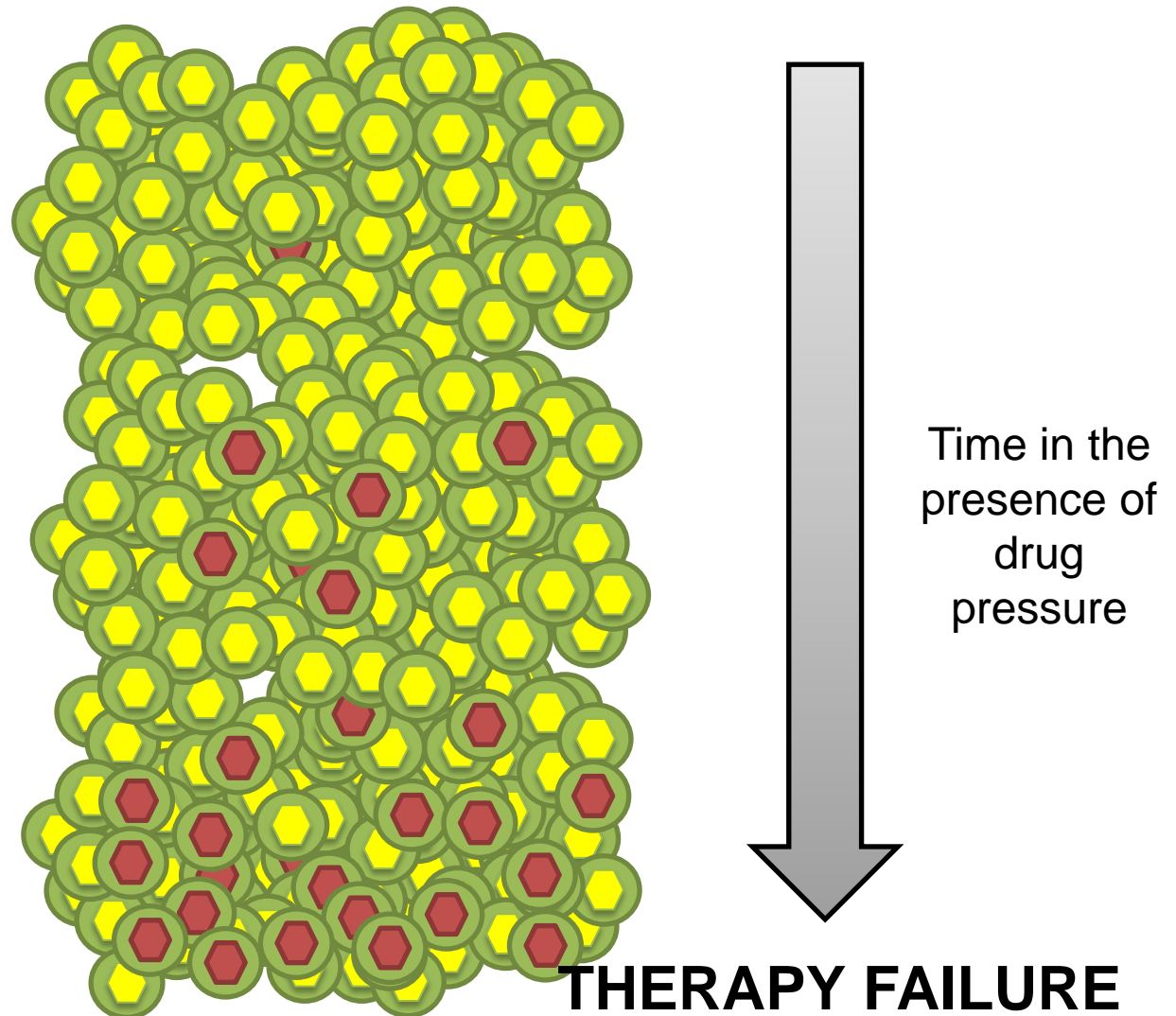
Sensitive

- Can detect drug resistance at a 0.1% of a patient's HIV virus population



- Laborious technique
- Research use only
- High cost per sample

Why is sensitive resistance testing important?



Low Frequency Resistance in VOICE & ASPIRE

VOICE	ASPIRE
<p>No data on low frequency NRTI resistance.</p> <p>Will seroconverters from tenofovir gel or oral TDF/FTC arms have low frequency resistance? Will it affect future first line treatment with Truvada?</p>	<p>Nevirapine (NVP)-resistant mutant frequencies >1% are significantly associated with increased risk of NVP-containing ART failure (A5208/Octane).</p> <p>Will low frequency NNRTI resistance affect efficacy of dapivirine ring? Will it affect future PMTCT or first line treatment with NVP or efavirenz?</p>

Resistance Testing at Virology Core

Receive samples

- Plasma samples received from sites
- QC on all shipments
- Log samples and assign testing

Endpoint confirmation

- EIA
- WB

Standard resistance testing

- Standard resistance testing (IHG) on all HIV positive samples with VL>200
- Send results to SCHARP and to sites

Sensitive resistance testing

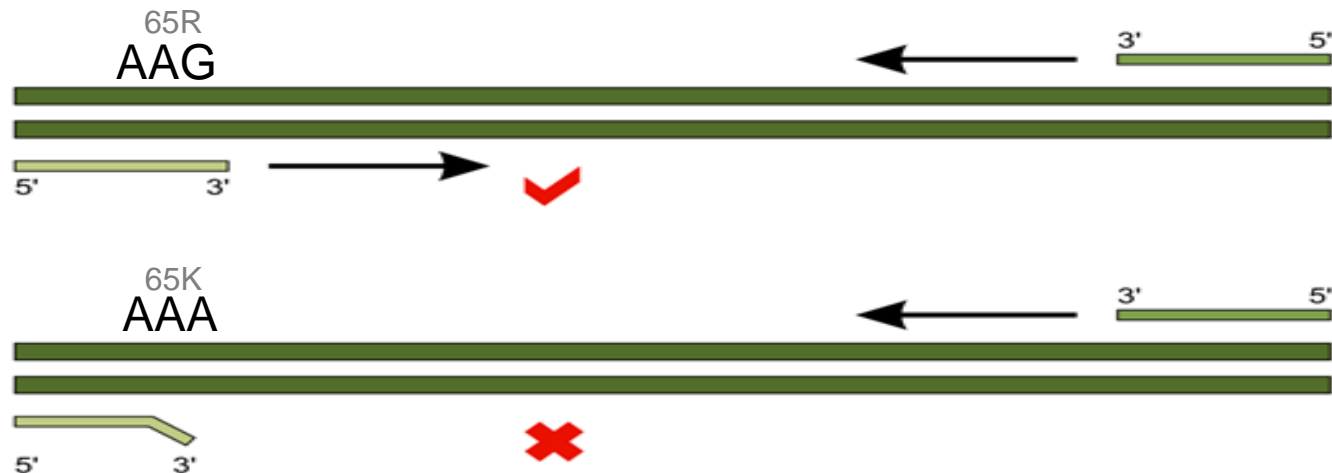
- Sensitive resistance testing (ASPCR or NGS) on all samples with successful IHG
- Send results to SCHARP

Standard vs Sensitive Tests

IHG In-House Genotyping	ASPCR Allele-Specific PCR	NGS Next-Generation Sequencing
<p>“Population” sequence</p> <p>Get one sequence that is the “consensus” for all viruses in that sample</p> <p>Can have mixed bases</p>	<p>Codon specific testing</p> <p>Provides frequency of wild-type vs mutant codon</p>	<p>Similar to IHG, except that “consensus” is not given as output. Individual sequences are generated for all HIV molecules amplified in each sample</p> <p>Can accurately quantitate mixtures at low frequency (0.1%)</p>

ASPCR

- Targets specific codons of interest
- Real-time PCR assay identifies % of viral templates with a specific codon



ASPCR Method

Step 1

- Extract HIV-1 RNA
- Convert RNA to cDNA

Step 2

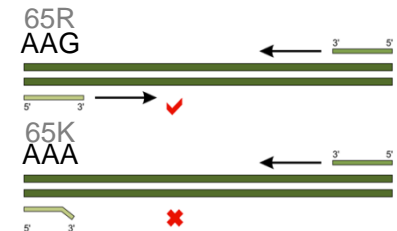
- Create large pool of templates through PCR amplification of patient HIV-1cDNA

Step 3

- Use ASPCR codon specific primers to determine presence of wild-type and mutant codons
- All samples are run with both primer sets

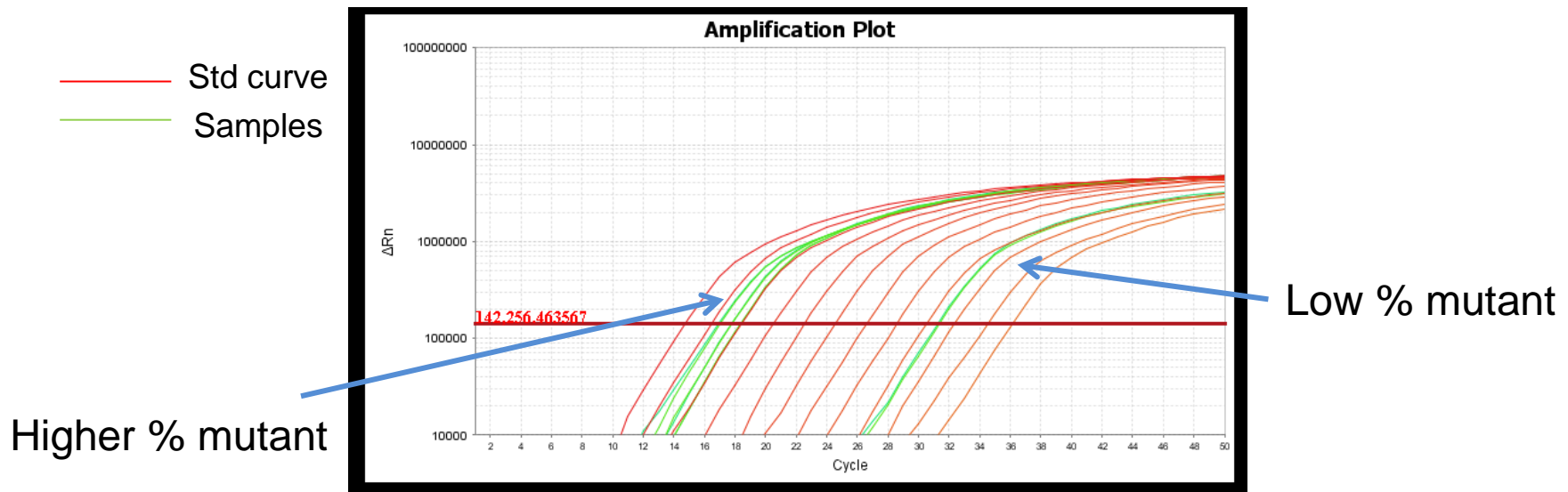
Output

- Presence and frequency of wild-type vs mutant codon is generated based on standard curves of wild-type and mutant mixtures



ASPCR results

- SYBR green based assay
- Frequency is determined by standard of wild-type and mutant mixtures of known %



VOICE ASPCR Results

Mutation	# Detected by Standard Genotyping/ # Seroconverted on Study Product	# Detected by ASPCR/ # Tested	Detection Limit	Range of Mutant Frequency
K65R	0/301	3/276	0.1%	0.5 – 15%
M184V	1/301	2/288	0.1%	0.5% - 98%
M184I	0/301	11/285	0.1%	0.5 – 5.2%
K70E	0/301	0/283	0.3%	-

- Detection of low frequency mutants did not differ across treatment arms or with the detection of tenofovir at any follow-up visit.
- Results presented at CROI 2015 (Panousis, et al.)

Sensitive Resistance Testing: VOICE

- **Resistance selection in VOICE remains LOW.**
- Mutant detection was **not** associated with treatment arm or detectable TFV.
 - Low frequency mutants may be transmitted resistance or spontaneously arising mutants of unknown clinical significance.
- Low product use in the VOICE trial could explain the infrequent selection of resistance among seroconverters.

NGS Method

Step 1

- Extract HIV-1 RNA
- Convert RNA to cDNA

Step 2

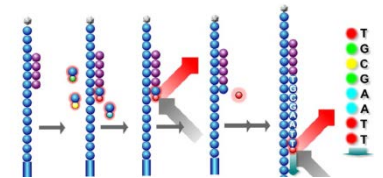
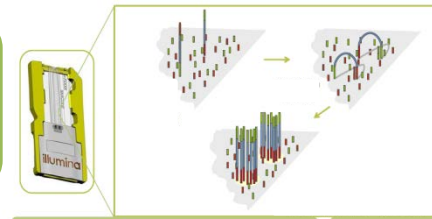
- Create large pool of templates through PCR amplification of patient HIV-1 cDNA

Step 3

- Use Illumina MiSeq platform to perform sequencing reaction

Output

- Sequence is generated for ALL amplified cDNAs
- Sequence read is the entire length of amplicon
- **100,000s of reads per sample!**



NGS results

- Output of assay= FASTQ files (like FASTA- string of sequence)



- Strong need for bioinformatics tools to process data
- Bioinformatics tools separate and analyze samples based on sample ID tags; report frequency of mutations in each sample

Next Steps

- Finish developing NGS assay for all subtypes
- Test VOICE samples with NGS to confirm low frequency mutations observed with ASPCR
- Test ASPIRE samples with NGS

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SCHARP

